



## Relationship Between Hormonal Variables and Bone Mineral Density, Muscle Force, and Fat Mass in Peripubertal Girls

Ivana Zofkova<sup>1\*</sup>, Veronika Cirmanova<sup>1</sup>, Petr Kasalicky<sup>2</sup>, Vera Lanska<sup>3</sup>, Vaclav Vyskocil<sup>4</sup>, Petr Matucha<sup>1</sup>, Milan Bayer<sup>5</sup>

<sup>1</sup>Institute of Endocrinology, Prague, Czech Republic

<sup>2</sup>Mediscan Group, Prague, Czech Republic

<sup>3</sup>Institute of Clinical and Experimental Medicine, Prague, Czech Republic

<sup>4</sup>Centrum of Osteology, Medical Faculty, Charles University, Pilsen, Czech Republic

<sup>5</sup>Department of Paediatrics, Charles University, Hradec Kralove Teaching Hospital, Prague, Czech Republic

### ARTICLE INFO

#### Article type:

Original Article

#### Article history:

Received: 30 Jun 2011

Revised: 02 Mar 2011

Accepted: 28 Mar 2011

#### Keywords:

Puberty

Insulin-Like Growth Factor I

Leptin

Estrogens

Serotonin

### ABSTRACT

**Background:** The muscle-bone unit represents an evolutionary system, in which both of its components are under the common control of the insulin-like growth factor I (IGF-I), sex hormones, and vitamin D. The mutual interactions between these hormones maintain integrity, growth and maturation of pubertal bone mass. Thus, insufficiency of any of these hormones will negatively influence development of the skeleton during puberty.

**Objectives:** The aim of the study as to analyse the correlation between muscle mass, total bone mineral content (BMC), bone mineral density (BMD) of the lumbar spine (BMD L<sub>1</sub>-L<sub>4</sub>), and serum or urine hormones.

**Materials and Methods:** Total BMC (g) and areal BMD L<sub>1</sub>-L<sub>4</sub> (g/cm<sup>2</sup> and Z-score) as well as muscle mass and fat mass (g) were assessed by means of dual-energy X-ray absorptiometry (DXA). The Z-score is the number of standard deviations a patient's BMD which differs from the average BMD of their age, sex, and ethnicity. This Parameter is used in children. Muscle force (N) was measured using a dynamometer.

**Results:** The simple correlations showed strong positive associations between BMC or BMD L<sub>1</sub>-L<sub>4</sub> (g/cm<sup>2</sup>) and serum phosphate, estradiol, insulin-like growth factor (IGF-I), leptin and fat masses, and muscle force ( $P < 0.001$  for all parameters). Positive correlations were also observed between BMD and serum phosphate ( $P < 0.01$ ), IGF-I ( $P < 0.01$ ), estradiol ( $P < 0.001$ ), leptin ( $P < 0.01$ ), fat and lean mass ( $P < 0.001$  and  $P < 0.001$ , respectively) and muscle force ( $P < 0.001$ ). The partial correlations, after eliminating the impact of height, Tanner stage, and physical activity level, confirmed positive relationships between either BMC or BMD L<sub>1</sub>-L<sub>4</sub> and lean mass ( $P < 0.001$  and  $P < 0.001$ , respectively) and fat mass ( $P < 0.001$  for BMC and BMD). Furthermore, a positive relationship was observed between serum leptin and both BMC and BMD (Z score) ( $P < 0.05$  and  $P < 0.05$ , respectively). After removing the effects of height, Tanner stage, and physical activity, positive associations were observed between lean mass and IGF-I ( $P < 0.01$ ), leptin levels ( $P < 0.05$ ), and muscle force ( $P < 0.01$ ).

**Conclusions:** On the basis of the study results, it can be expected that low values of lean or fat mass, and insufficient production of IGF-I or leptin, could negatively influence bone development in pubertal girls.

Copyright © 2011 Kowsar M. P. Co. All rights reserved.

#### ► Implication for health policy/practice/research/medical education:

Monitoring of bone mineral density together with muscle indices and hormonal parameters in pubertal girls could prevent development of osteoporosis in women.

\* Corresponding author: Ivana Zofkova, Institute of Endocrinology, Narodni 8, 116 94 Prague 1. Tel: +420-224905412, Fax: +420-22490325, E-mail: izofkova@endo.cz

DOI: 10.5812/Kowsar.1726913X.2583

Copyright ©2011 Kowsar M.P.Co. All rights reserved.

► Please cite this paper as:

Zofkova I, Cirmanova V, Kasalichy P, Lanska V, Vyskocil V, Matucha P, et al. Relationship Between Hormonal Variables and Bone Mineral Density, Muscle Force, and Fat Mass in Peripubertal Girls. *Int J Endocrinol Metab.* 2011;9(3): 391-6. DOI: 10.5812/Kowsar.1726913X.2583

## 1. Background

A bone is not only a motor apparatus, but also a living tissue, both producing and receiving many hormones. The effects of estrogens, androgens, growth factors, the active metabolite of vitamin D, and many other hormones on bone remodelling are largely related to receptor activator of nuclear factor- $\kappa$ B ligand/receptor activator of nuclear factor- $\kappa$ B/osteoprotegerin (RANKL/RANK/OPG system) modulation. This signalling loop directly regulates the activity of osteoclasts and osteoblasts and mediates the effects of hormones and cytokines on the skeleton (1). The concept of the muscle-bone unit assumes that bones and muscles represent an evolutionary functional unit, and both of its components are under the control of the insulin-like growth factor I (IGF-I), sex hormones and vitamin D (2,3). IGF-I has a direct anabolic effect on bone (4). In mutual interactions with oestrogens and androgens (5) this peptide initiates growth spurts and accelerates maturation of bone mass during puberty and adolescence (6). The highest levels of IGF-I are reported between the 3<sup>rd</sup> and 4<sup>th</sup> Tanner stage and corresponding to bone maturity (7). It is expected that variations in IGF-I expression are one of the causes for inter individual differences in peak bone mass values, its quality and dimension of the skeleton in humans with a positive energy balance and adequate physical activity (8).

The next hormone regulating bone development is vitamin D. It was observed that more serious cases of hypovitaminosis D slowed muscle and bone development with the effect being significantly more prevalent in girls than in boys, particularly when associated with low levels of physical activity (9). However, the relationship between D vitamin homeostasis and the muscle-bone unit in peripubertal girls has not been extensively studied, despite a generally high prevalence of hypovitaminosis D in children and adolescents. Inconsistent data have been obtained on the role of adipokines and the serotonin - melatonin axis in bone development. Leptin activates the receptors of osteoblasts and thus, directly increases bone mass accrual. However, leptin also exerts a central inhibitory effect on the skeleton via  $\beta_2$  receptors, which activate bone turnover (10).

In recent years, attention has been focused on osteotropic effects of neurotransmitter 5-hydroxytryptamine (serotonin). Dampening of serotonin production, induced by Wnt/Lrp/ $\beta$ -catenin circuit activation, has an anabolic effect on bones (11). Furthermore, higher serotonin levels could explain an increased incidence of fractures in patients treated with antidepressants that inhibit serotonin reuptake (12). The relevance of melatonin in pubertal bone development is not clear. It is known, that

this indole inhibits production of gonadotropins (13). Conversely, it has direct anabolic effects on bones, mediated by specific receptors on osteoblasts that induce inhibition of adipocytogenic differentiation and activation of osteogenesis (14-18). The authors are unaware of any specific investigations regarding the role of the serotonin - melatonin system in the regulation of bone development.

## 2. Objectives

The goal of this study was to assess the relationships between hormones, soft tissues, and the total bone mineral content (BMC) (g) or bone mass density (BMD) of the L<sub>1</sub>-L<sub>4</sub> vertebrae (g/cm<sup>2</sup> or Z-score) in a group of healthy peripubertal girls.

## 3. Materials and Methods

### 3.1. Subjects

A total of 100 healthy girls, aged 9-15 years (average 12.3  $\pm$  1.4), randomly selected from several regions in Prague, were examined. Inclusion criteria comprised the following: having reached puberty stage 1-4, having a good nutritional state, and consenting to follow the terms of the study protocol. The stage of puberty was determined according to an assessment of the extent of axial hair (Tanner A), pubic hair (Tanner P) and breast development (Tanner M). Thirty girls were assessed at Tanner 1, thirty-six girls at Tanner 2, ten girls at Tanner 3, and twenty-four girls were in Tanner 4 stage. Forty-two of our girls were menarcheal. Exclusion criteria were as follows: malnutrition, milk intolerance, any internal or psychiatric diseases, or current treatment with medications. Information about physical activity (evaluated as: grade 1, for normal school exercise once a week, grade 2, for regular participation in recreational sports, and grade 3, for those involved in racing sports), dietary habits (intake of dairy products), and neonatal data (fetal maturity at birth and duration of breast feeding) were gained from their parents through the use of a questionnaire. Informed consent was obtained from all of these girls and their parents and all procedures were approved by the Ethics Committee of the Institute of Endocrinology in Prague.

### 3.2. Protocol

Blood samples were taken from fasting girls in the morning (between 7 and 8 a.m.) to assess estradiol, follicle-stimulating hormone (FSH), luteinizing hormone (LH), vitamin 25(OH)D, parathyroid hormone (PTH), IGF-I, leptin, adiponectin, osteocalcin, bone isoenzyme of alkaline phosphatase (bALP), and total calcium and phosphorus. In all girls nocturnal melatonin

and serotonin excretion was determined in urine collected from 9 p.m. to 6 a.m. To minimize the effect of estradiol fluctuations during the menstrual cycle, samples from the 42 girls, who had attained menarche was performed immediately after menstruation.

In all girls, values of total BMC ( $\text{g}/\text{cm}^2$ ), BMD  $L_1-L_4$  ( $\text{g}/\text{cm}^2$  and its Z-score (referenced to a healthy population of the same age), lean (muscle) and fat mass were measured by use of dual-energy X-ray absorptiometry (DXA). The precision of the method is 1%. Muscle force was measured with a dynamometer (N) and was expressed as the sum of isometric flexion and extension of fingers, arms, lower legs, and the flexion and extension of the trunk.

### 3.3. Laboratory Studies

Estradiol was estimated using a commercial kit from the HUMA LAB CS. The inter-assay coefficient of varia-

tion (CV) was 15%. FSH and LH were measured using HUMA LAB CS kits (Slovak Republic). 25(OH) vitamin D was measured using an RIA kit (Immunodiagnostic System Holdings PLC) (normal range 9.2 - 45.2 ng/mL, CV 9.9%). Serum PTH was determined using electrochemiluminescence (ECLIA) assay (upper normal limit 65 ng/mL, CV 6.5%). Serum leptin was assessed using an RIA kit (LINCO Research, (St. Charles, MO, USA) (normal range in women 7.4-3.7 ng/mL, CV 4.0%), adiponectin using an RIA kit from LINCO Research (CV 6.9%), serum IGF-I and IGFBP-3 using IRMA methods from IMMUNOTECH (Marseille, France) (normal values of IGF-I in healthy children 114-400 ng/mL depending on stage of puberty, CV 6.8%). Nocturnal urinary melatonin sulphate excretion was measured using an ELISA immunoassay, IBL (Hamburg, Germany) (mean value in healthy adolescents 2.8  $\mu\text{g}/\text{h}$ ), and a similar method was used for the determination of nocturnal urinary serotonin excretion (normal values

**Table 1.** Anthropometric and Biochemical Characteristics of the Study Group

	MV	SD	MIN	MED	MAX
Age, y	12.299	1.362	9.267	12.313	15.358
Weight, kg	44.999	11.209	22	44.5	67
Height, cm	153.96	11.226	126.5	150.15	173.8
BMI, $\text{kg}/\text{m}^2$	18.791	3.521	12.252	19.56	27.76
Tanner M	2.282	1.151	1	2.5	4
Tanner P	2.212	1.156	1	2.5	4
Tanner A	2.224	1.127	1	2.5	4
Physical activity	2.012	0.681	1	2	3
Muscle force, N	2430	47.8	1253	2502	3750
Gestation, Wk	39.635	1.022	33	37.5	42
Breast feeding, mo	6.282	4.734	0	12	24
Fat, g	13114	6982.5	2064	15404.5	28805
Lean, g	31236	5907.3	20449	36723	42997
BMC total, g	1749.1	456.37	953.6	1810.8	2688
BMD $L_1-L_4$ , $\text{g}/\text{cm}^2$	0.933	0.176	0.643	1.006	1.369
BMD Z-score	0.171	1.025	-2,5	-0.15	2.2
Serum					
Osteocalcin, $\mu\text{g}/\text{L}$	143.4	48.643	48	148.9	249.8
bALP, $\mu\text{g}/\text{L}$	81.376	32.448	15.03	84.365	153.7
Ca, $\text{mmol}/\text{L}$	2.475	0.143	2.18	2.515	2.85
P, $\text{mol}/\text{L}$	1.463	0.161	1.14	1.5	1.86
25(OH)D, $\mu\text{g}/\text{L}$	18.985	6.901	10.09	28.945	47.8
PTH, $\text{ng}/\text{L}$	42.744	12.677	18.5	47.3	76.1
IGF-I, $\text{ng}/\text{mL}$	664.72	236.25	240.5	711.3	1182.1
Estradiol, $\text{nmol}/\text{L}$	0.204	0.191	0.007	0.487	0.967
adiponectin, $\text{ng}/\text{mL}$	13.554	4.983	5.71	18.58	31.45
Leptin, $\text{ng}/\text{mL}$	4.952	2.691	0.85	6.04	11.23
Urine					
Melatonin, $\mu\text{g}/\text{hour}$	107.31	106.89	11.4	294.25	577.1
Serotonin, $\mu\text{g}/\text{night}$	131.43	179.14	0.21	470.06	939.9

**Table 2.** Partial Correlations Between Bone and Body Composition, Muscle force, Physical Activity, and Hormones.

	Osteocalcin	bALP	BMC Total	BMD L <sub>1</sub> -L <sub>4</sub>	BMD Z- Score
Calcium	Calcium	0.008	-0.099	-0.149	-0.080
Phosphate	Phosphate	0.353 <sup>c</sup>	-0.379 <sup>c</sup>	-0.414 <sup>c</sup>	-0.273 <sup>b</sup>
25(OH)D	25(OH)D	-0.063	-0.120	-0.016	0.125
1,25(OH) <sub>2</sub> D <sub>3</sub>	1,25(OH) <sub>2</sub> D <sub>3</sub>	0.124	-0.056	0.041	0.040
PTH	PTH	0.133	0.029	0.025	-0.159
IGF-I	IGF-I	0.185 <sup>a</sup>	0.450 <sup>c</sup>	0.489 <sup>c</sup>	0.304 <sup>b</sup>
IGF-IBP3	IGF-IBP3	-0.153	0.463 <sup>c</sup>	0.463 <sup>c</sup>	0.228
Estradiol	Estradiol	-0.449 <sup>c</sup>	0.487 <sup>c</sup>	0.543 <sup>c</sup>	0.319 <sup>c</sup>
Adiponectin	Adiponectin	0.083	-0.176	-0.107	-0.253
Leptin	Leptin	-0.155	0.350 <sup>c</sup>	0.331 <sup>c</sup>	0.288 <sup>b</sup>
Melatonin	Melatonin	0.017	-0.114	-0.064	-0.166
Serotonin	Serotonin	-0.024	-0.001	0.040	-0.019
Fat mass	Fat mass	-0.332 <sup>c</sup>	0.581 <sup>c</sup>	0.499 <sup>c</sup>	0.527 <sup>c</sup>
Lean mass	Lean mass	-0.380 <sup>c</sup>	0.898 <sup>c</sup>	0.809 <sup>c</sup>	0.554 <sup>c</sup>
Muscle force	Muscle force	-0.260 <sup>b</sup>	0.693 <sup>c</sup>	0.643 <sup>c</sup>	0.426 <sup>c</sup>
Physical activity	Physical activity	-0.020	0.165	0.275 <sup>b</sup>	0.259 <sup>a</sup>
Breast feeding	Breast feeding	-0.002	-0.002	-0.020	0.118
Neonatal maturity	Neonatal maturity	-0.020	0.003	0.016	0.147

<sup>a</sup>  $P < 0.05$ , <sup>b</sup>  $P < 0.01$ , <sup>c</sup>  $P < 0.001$ 

were not determined). Total blood calcium and phosphate were measured using absorption spectrophotometry. Serum osteocalcin was determined using an ECLIA method from Roche Diagnostics (Mannheim, Germany) and bone alkaline phosphatase (bALP) isoenzyme analysed using an IRMA kit from IMMUNOTECH (a Beckman Coulter company).

### 3.4. Statistics

The mean, standard deviation, median, maximum and minimum values were used for description of the investigated variables. To measure the dependence of two variables, while the subgroup of chosen variables was held constant, the partial correlation coefficient was used. Normality was tested by the Shapiro-Wilk's *W* statistic. All significant tests were two-sided and  $P < 0.05$  as regarded as significant. Statistical software (SYSTAT Software, Inc., Chicago, IL, USA) was used for calculations.

## 4. Results

Table 1 shows the mean values ( $\pm$  SD) of all analysed parameters, including anthropometric data and the stage of puberty (Tanner 1-4).

Table 2 shows the unadjusted partial correlations between bone and body composition, muscle force, physical activity, and hormones. The simple correlations showed strong positive associations between either BMC or BMD L<sub>1</sub>-L<sub>4</sub> and serum phosphate, estradiol, IGF-I, leptin, lean and fat volumes, and muscle force ( $P < 0.001$ ). Posi-

tive correlations were observed between BMD L<sub>1</sub>-L<sub>4</sub> and serum phosphate ( $P < 0.01$ ), IGF-I ( $P < 0.01$ ), estradiol ( $P < 0.001$ ), leptin ( $P < 0.01$ ), fat and lean masses ( $P < 0.001$  for both the latter indices), and muscle force ( $P < 0.001$ ) after removing linear effects of height, Tanner stage, and physical activity.

Table 3 indicates partial correlations of osteocalcin, bALP, BMC total, BMD L<sub>1</sub>-L<sub>4</sub> and BMD Z-score with calcium, phosphate, serum and urine hormones, soft tissues, and neonatal parameters. Strong positive correlations were found between lean mass and BMC or BMD Z-score ( $P < 0.001$  and  $P < 0.001$ , respectively), as well as between fat and BMC, BMD L<sub>1</sub>-L<sub>4</sub> (g/cm<sup>2</sup>) and BMD (Z-score) ( $P < 0.001$ ,  $P < 0.01$  and  $P < 0.001$ ). A positive association was observed between leptin and the BMC and BMD Z-score ( $P < 0.05$  for both bone parameters). Positive relationships were observed between serum osteocalcin and serum phosphate, 25 (OH) vitamin D and urine melatonin ( $P < 0.01$ ,  $P < 0.05$ , and  $P < 0.05$ , respectively). Similarly a positive correlation was seen between urine serotonin and bALP ( $P < 0.05$ ). However, none of these hormonal parameters was correlated with BMC or BMD L<sub>1</sub>-L<sub>4</sub>. No correlations were observed between bone parameters and serum adiponectin. Partial correlations between lean mass or muscle force and hormone indices, after removing linear effects of height, Tanner stage and physical activity, are depicted in Table 4, which shows positive associations between lean volume or muscle force and serum IGF-I ( $P < 0.01$  and  $P < 0.01$ , respectively) and between lean volume (but not muscle force) and serum leptin ( $P < 0.05$ ). No as-

**Table 3.** Partial Correlations of Osteocalcin, bALP, BMC Total, BMD with Calcium, Phosphate, Serum and Urine Hormones, Soft Tissues, and Neonatal Parameters after Removing the Linear Effects of Height, Tanner Stage, and Physical Activity

	Osteocalcin	bALP	BMC Total	BMD L1-L4	BMD Z-Score
Ca	0.112	-0.061	0.022	0.004	-0.018
P	0.299 <sup>b</sup>	-0.028	-0.032	-0.093	-0.017
25(OH)D	0.252 <sup>a</sup>	-0.089	0.015	-0.001	0.028
PTH	0.227 <sup>a</sup>	0.184	-0.096	-0.111	-0.122
IGF-I	0.174	-0.007	0.090	0.074	0.131
IGF-I/IGFBP3	-0.069	-0.152	0.148	0.103	0.060
Estradiol	-0.037	-0.303 <sup>b</sup>	0.142	0.143	-0.096
Adiponectin	-0.018	0.107	0.211	-0.189	-0.252
Leptin	-0.214	0.115	0.303 <sup>a</sup>	0.217	0.275 <sup>a</sup>
Melatonin	0.313 <sup>b</sup>	-0.122	-0.138	-0.130	-0.242
Serotonin	0.168	0.234 <sup>a</sup>	-0.232	-0.245	-0.052
Fat mass	-0.385 <sup>b</sup>	-0.039	0.564 <sup>c</sup>	0.377 <sup>b</sup>	0.482 <sup>c</sup>
Lean mass	-0.100	-0.004	0.666 <sup>c</sup>	0.188	0.551 <sup>c</sup>
Muscle force	-0.046	0.001	0.349 <sup>b</sup>	0.188	0.242
Gestation	0.125	0.120	0.196	0.194	0.190
Breast feeding	-0.029	-0.002	0.011	-0.035	-0.061

<sup>a</sup>  $P < 0.05$ , <sup>b</sup>  $P < 0.01$ , <sup>c</sup>  $P < 0.001$

**Table 4.** Partial Correlations of Lean Mass and Muscle Force with Calcium, Phosphate, and Serum and Urine Hormones after Removing the Effects of Height, Tanner Stage, and Physical Activity.

	Lean Mass	Muscle Force
Ca	-0.024	-0.013
P	0.163	-0.021
25(OH)D	0.009	-0.212
1,25(OH)2D3	0.049	-0.050
PTH	0.118	0.034
IGF-I	0.363 <sup>b</sup>	0.261 <sup>b</sup>
IGFII/IGFBP3	0.302	0.308
Estradiol	0.043	-0.145
Adiponectin	-0.120	-0.228 <sup>a</sup>
Leptin	0.311 <sup>a</sup>	0.105
Melatonin	-0.046	0.039
Serotonin	-0.067	-0.169

<sup>a</sup>  $P < 0.05$ , <sup>b</sup>  $P < 0.01$

sociations were found between the neonatal parameters and BMC or BMD.

## 5. Discussion

The study confirmed a positive association between pubertal bone and fat, or its hormone leptin, which is in agreement with the study on pubertal girls of Rhie *et*

*al.* (19). Thus, both these studies support the hypothesis that the direct stimulating effect of leptin on developing bone mass prevails over its inhibitory effect mediated through central mechanisms (10).

In our girls the positive relationship was also found between leptin and lean mass values, which corresponds with the interesting results obtained by Olmedillas *et al.*, who found up-regulation of leptin receptors in the hypertrophic biceps of professional tennis players [24]. These data allow us to hypothesize that the positive effect of leptin on bone is partially mediated by muscle. It is generally accepted that muscle function is positively influenced by vitamin D. Ward *et al.* (9) demonstrated a close positive correlation between 25(OH) vitamin D levels and both muscle strength and high jump results in adolescent girls, with a corresponding negative relationship between PTH levels and these parameters. Moreover, positive correlations between 25(OH) vitamin D and vertebral BMD and trochanter BMC has been documented in the study performed by El-Hajj Fuleihan *et al.* (20). In our study, however, we were unable to demonstrate any association between serum 25(OH) vitamin D or PTH and either lean or bone mass. The reason for the lack of an obvious relationship between 25(OH) vitamin D levels and muscle function is not clear from our study, but could be partially explained by the fact that girls in the El-Hajj Fuleihan study were undernourished and the girls studied by Ward were younger than our participants. The start of female puberty is induced by an increase in sexual steroid production. The osteotropic effect of estrogens (or androgens) is direct, but partly mediated by IGF-I provided that the nutritional status and physical activity levels are appropriate (21). In our study relationships between bone parameters and estradiol or IGF-I levels could be showed in unadjusted analyses. However, these associations lost statistical significance after eliminating the effects of age, Tanner stage, and physical activity level. Since IGF-I production and bone response to the hormone depend partly on physical activity (7), the lack of an adjusted-association between serum IGF-I levels and bone mass could be explained by relatively low physical activity levels in our girls. On the other hand, the effect of protein-caloric malnutrition could be excluded for the majority of our participants. Serotonin has a bimodal influence on target tissues, including bones, depending on the source of its synthesis (duodenum or central nervous system). Serotonin produced by enterochromaffin cells (95% of total production) inhibits bone formation, whereas serotonin produced in the central nervous system has the opposite effect (22). The inconclusiveness of association between serotonin levels and bone parameters in our girls could be explained by the sum of impacts from both of these serotonin pools. The concept of osteoanabolic effects for melatonin is supported in the present study by a proven positive association between nocturnal melatonin excretion levels and osteocalcin levels. However, we did not observe any relationships be-

tween serum melatonin and either BMC or BMD, which could be, in part, explained by the physiological decline of this hormone during puberty. Thus, melatonin does not seem to play a key role in the development of bones in human females (23).

As expected, the study demonstrated significant positive associations between bone mass and serum leptin levels in peripubertal girls. However, the study yielded some surprising results. It failed to demonstrate any relationship between bone parameters and IGF-I levels, even though IGF-I is positively correlated with muscle mass and muscle force. Similarly, the association between muscle parameters and 25 (OH) vitamin D levels was inconclusive in girls. Nevertheless, the study allows us to theorize that a delay in puberty and the deficient development of muscle or fat slows bone mass development via its direct effects on bones, independently of the vitamin D status. Positive associations between the factors of lean tissue mass, muscle force, and physical activity on the one hand and BMC or BMD on the other hand are compatible with the muscle-bone unit concept. Continuous monitoring of sexual development, together with tissue composition and hormone parameters (leptin and IGF-I), might help in the early identification of girls with a later high risk of fractures due to inadequate development of peak bone mass.

## Acknowledgments

None declared.

## Financial Disclosure

None declared.

## Funding/Support

None declared.

## References

- Ostrowska Z. [Menopause, obesity, and bone status]. *Postepy Hig Med Dosw (Online)*. 2009;**63**:39-46.
- Ashby RL, Adams JE, Roberts SA, Mughal MZ, Ward KA. The muscle-bone unit of peripheral and central skeletal sites in children and young adults. *Osteoporos Int*. 2011;**22**(1):121-32.
- Fricke O, Beccard R, Semler O, Schoenau E. Analyses of muscular mass and function: the impact on bone mineral density and peak muscle mass. *Pediatric Nephrology*. 2010;**25**(12):2393-400.
- Mohan S, Baylink DJ. Impaired skeletal growth in mice with haploinsufficiency of IGF-I: genetic evidence that differences in IGF-I expression could contribute to peak bone mineral density differences. *J Endocrinol*. 2005;**185**(3):415-20.
- Venken K, Schuit F, Van Lommel L, Tsukamoto K, Kopchick JJ, Coschigano K, et al. Growth without growth hormone receptor: estradiol is a major growth hormone-independent regulator of hepatic IGF-I synthesis. *J Bone Miner Res*. 2005;**20**(12):2138-49.
- Bonjour JP, Chevalley T, Ferrari S, Rizzoli R. The importance and relevance of peak bone mass in the prevalence of osteoporosis. *Salud Publica Mex*. 2009;**51** (Suppl 1):S5-17.
- Maimoun L, Coste O, Galtier F, Mura T, Mariano-Goulart D, Paris F, et al. Bone mineral density acquisition in peripubertal female rhythmic gymnasts is directly associated with plasma IGF-I/IGF-binding protein 3 ratio. *Eur J Endocrinol*. 2010;**163**(1):157-64.
- Karatay S, Yildirim K, Melikoglu M, Akcay F, Şenel K. Effects of dynamic exercise on circulating IGF-I and IGFBP-3 levels in patients with rheumatoid arthritis or ankylosing spondylitis. *Clin Rheumatol*. 2007;**26**(10):1635-9.
- Ward KA, Das G, Berry JL, Roberts SA, Rawer R, Adams JE, et al. Vitamin D status and muscle function in post-menarchal adolescent girls. *J Clin Endocrinol Metab*. 2009;**94**(2):559-63.
- Hipmair G, Böhler N, Maschek W, Soriguer F, Rojo-Martinez G, Schimetta W, et al. Serum leptin is correlated to high turnover in osteoporosis. *Neuro Endocrinol Lett*. 2010;**31**(1):155-60.
- Yadav VK, Balaji S, Suresh PS, Liu XS, Lu X, Li Z, et al. Pharmacological inhibition of gut-derived serotonin synthesis is a potential bone anabolic treatment for osteoporosis. *Nat Med*. 2010;**16**(3):308-12.
- Warden SJ, Hassett SM, Bond JL, Rydberg J, Grogg JD, Hilles EL, et al. Psychotropic drugs have contrasting skeletal effects that are independent of their effects on physical activity levels. *Bone*. 2010;**46**(4):985-92.
- Srinivasan V, Spence WD, Pandi-Perumal SR, Zakharia R, Bhatnagar KP, Brzezinski A. Melatonin and human reproduction: shedding light on the darkness hormone. *Gynecol Endocrinol*. 2009;**25**(12):779-85.
- Radio NM, Doctor JS, Witt-Enderby PA. Melatonin enhances alkaline phosphatase activity in differentiating human adult mesenchymal stem cells grown in osteogenic medium via MT2 melatonin receptors and the MEK/ERK (1/2) signaling cascade. *J Pineal Res*. 2006;**40**(4):332-42.
- Sanchez-Hidalgo M, Lu Z, Tan D-X, Maldonado MD, Reiter RJ, Gregerman RL. Melatonin inhibits fatty acid-induced triglyceride accumulation in ROS17/2.8 cells: implications for osteoblast differentiation and osteoporosis. *Am. J. Physiol. Regul. Integr. Comp. Physiol*. 2007;**292**(6):R2208-R15.
- Suzuki N, Somei M, Seki A, Reiter RJ, Hattori A. Novel bromomelatonin derivatives as potentially effective drugs to treat bone diseases. *J Pineal Res*. 2008;**45**(3):229-34.
- Witt-Enderby PA, Radio NM, Doctor JS, Davis VL. Therapeutic treatments potentially mediated by melatonin receptors: potential clinical uses in the prevention of osteoporosis, cancer and as an adjuvant therapy. *J Pineal Res*. 2006;**41**(4):297-305.
- Zhang L, Su P, Xu C, Chen C, Liang A, Du K, et al. Melatonin inhibits adipogenesis and enhances osteogenesis of human mesenchymal stem cells by suppressing PPARgamma expression and enhancing Runx2 expression. *J Pineal Res*. 2010;**49**(4):364-72.
- Rhie YJ, Lee KH, Chung SC, Kim HS, Kim DH. Effects of body composition, leptin, and adiponectin on bone mineral density in prepubertal girls. *J Korean Med Sci*. 2010;**25**(8):1187-90.
- El-Hajj Fuleihan G, Nabulsi M, Tamim H, Maalouf J, Salamoun M, Khalife H, et al. Effect of vitamin D replacement on musculoskeletal parameters in school children: a randomized controlled trial. *J Clin Endocrinol Metab*. 2006;**91**(2):405-12.
- Jurimae J, Maestu J, Jurimae T. Bone turnover markers during pubertal development: relationships with growth factors and adipocytokines. *Med Sport Sci*. 2010;**55**:114-27.
- Ducy P, Karsenty G. The two faces of serotonin in bone biology. *J Cell Biol*. 2010;**191**(1):7-13.
- Molina-Carballo A, Fernandez-Tardaguila E, Uberos-Fernandez J, Seiquer I, Contreras-Chova F, Munoz-Hoyos A. Longitudinal study of the simultaneous secretion of melatonin and leptin during normal puberty. *Horm Res*. 2007;**68**(1):11-9.
- Olmedillas H, Sanchis-Moysi J, Fuentes T, Guadalupe-Grtao A, Ponce-González JG, Morales-Alamo D et al. Muscle hypertrophy and increased expression of leptin receptors in the mucusculus triiceps brachii of the dominant arm in professional tennis players. *Eur J Appl Physiol* 2010; **108** (4):749-58.