

# Study of the Antibacterial Activity of Methanolic and Aqueous Extracts of *Myrtus communis* on Pathogenic Strains Causing Infection

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## Abstract

**Background:** Medicine plants have been used as sources of medicine in virtually all cultures. During the last decade, the use of traditional medicine (TM) has been expanded globally and is gaining popularity.

**Objectives:** The antimicrobial activities of methanol and water extracts of *Myrtus communis* L. leaves were evaluated in this study.

**Materials and Methods:** In this experimental study, the tests were carried out using disk agar diffusion method at four extract concentrations (5, 10, 15 and 20 mg/mL). The MICs and MBCs of the extracts of *M. communis* were determined by agar dilution method. Average results were reported as the mean and standard error (mean  $\pm$  SE) and SPSS-18 statistical software, oneway ANOVA followed by Turkey's test were used to do inter-group comparison, while considering  $P \leq 0.05$  as the significance level.

**Results:** Methanol extract of *M. communis* exhibited significant antibacterial activity in the concentration of 20 mg/mL ( $P \leq 0.05$ ) against *Staphylococcus epidermidis* and *Enterococcus faecalis* with a greater inhibition zone of 20 mm, while a 14 mm zone of inhibition was observed in *Escherichia coli* and *Shigella flexneri*. The minimum inhibitory concentration (MIC) of the extracts ranged between 2 mg/mL and 128 mg/mL while the minimum bactericidal concentration (MBC) ranged between 4 mg/mL and 256 mg/mL.

**Conclusions:** The study showed that species, strains and concentrations of *M. communis* extract are of those factors that may influence the sensitivity of the tested bacteria. A significant correlation was observed between zone of inhibition and concentration of extract.

**Keywords:** Infection, Aqueous Extracts, Methanolic Extract, *Myrtus communis*

## 1. Background

The rediscovery of the connection between plants and health is responsible for the launching of new generations of multi-component botanical drugs, dietary supplements and plant-produced recombinant proteins [1]. Approximately 700 mono and poly-herbal preparations in the form of decoction, tincture, tablets and capsules from more than 100 plants are in clinical use [2].

Though the resistance development by microbes cannot be stopped, appropriate action will reduce the mortality and health care costs by using antibiotic resistant inhibitors of plant origin [3]. The increase in resistance of microorganisms due to multi-various use of commercial antimicrobial drugs has encouraged scientists all over the world to search for new antimicrobial substances from various antecedents comprising medicinal plants [4].

Myrtle (*Myrtus communis* L.) is a member of Mirtaceae family that grows spontaneously throughout the Mediterranean area [5]. The plant is an ever green shrub or small

tree, growing to 5 metres (16 ft) tall. The entire leaf is, 3 - 5 cm long, with a fragrant essential oil. The leaves contain tannins, flavonoids such as quercetin, catechin and myricetin derivatives and volatile oils Myrtle occupies a prominent place in the writings of Hippocrates, Pliny, Dioscorides, Galen, and the Arabian writers. In several countries, particularly in Europe and China, there has been a tradition for prescribing this substance for sinus infections [6].

Infections are associated with intravascular devices (prosthetic heart valves, shunts, etc.) and also commonly occur in prosthetic joints, catheters, and large wounds. Catheter infections along with catheter-induced UTIs lead to serious inflammation and pus secretion. In these instances, urination is extremely painful. Septicemia and endocarditis are also diseases associated with *Staphylococcus epidermidis* [7]. Their symptoms run the gamut from fever, headache and fatigue to anorexia and dyspnea. Septicemia is especially prevalent resulting from neonatal infections,

particularly in very low birth weights. Endocarditis is an infection of the heart valves and parts of the inside lining of the heart muscle. *S. epidermidis* is very likely to contaminate patient-care equipment and environmental surfaces, possibly explaining the high incidence of *S. epidermidis* in the hospital setting [8]. Several different serogroups of *Shigella* are described; *Shigella flexneri* belongs to group B. *S. flexneri* infections can usually be treated with antibiotics, although some strains have become resistant [9]. *Enterococcus faecalis* can and *Escherichia coli* cause endocarditis and bacteremia, urinary tract infections (UTI), meningitis, and other infections in humans [10].

## 2. Objectives

On the basis, the aim of this work was to investigate the possible inhibitory activity of *Myrtus communis* extracts against pathogenic strains causing infection.

## 3. Materials and Methods

This experimental study was conducted at industrial microbiology laboratory, department of food sciences and technology, Ferdowsi University of Mashhad in 2014.

### 3.1. Media

Mueller-Hinton agar and nutrient agar (DEFCO Laboratories, USA), peptone bacteriological (BDH Chemicals Ltd., England), tryptone soya broth and sodium chloride (East Anglia Chemicals, UK) were all obtained from the department of food sciences and technology, Ferdowsi University of Mashhad.

### 3.2. Test Strains

The following Gram (+) and Gram (-) bacteria were used for testing antibacterial activity. *S. epidermidis* ATCC 12228, *E. faecalis* ATCC 29212, *E. coli* ATCC29998 and *S. flexneri* ATCC12022 all American type culture collections were obtained from the microbiology department of Mashhad University of Medical Sciences, Mashhad.

### 3.3. Collection and Preparation of the Plant Material

The *Myrtus communis* were collected from countryside of Behbahan (Khuzestan Province, Iran). *M. communis* were dried in an open air protected from direct exposure to sunlight. The dried plant materials were separately powdered to suitable size [11]. The identity of each plant specimen was confirmed at the Herbarium, Ferdowsi University of Mashhad where a voucher specimen was deposited.

### 3.4. Plant Extraction

Two hundred grams of each powdered plant material were extracted with 80% methanol or water by maceration for 72 hours with frequent agitation and the resulting liquid was filtered (Whatman No. 3 filter paper, Whatman Ltd., England). Extraction was repeated five times and the filtrates of all portions were combined in one vessel. The organic solvent was removed by evaporation using Rota vapor (Rota-vapor R-205, England) at no more than 35°C. The aqueous residues were then placed in an oven at 35°C for about 72 hours to remove the water. The resulting dried mass was then powdered, packed into a glass vial and stored in a desiccator over silica gel until use [12].

### 3.5. Suspension Preparation

Fresh cultivated *S. epidermidis* ATCC 12228, *E. faecalis* ATCC 29212, *E. coli* ATCC29998 and *S. flexneri* ATCC12022 colonies were suspended in 10 mL of 0.80% normal saline. Suspension was mixed for 20 seconds with a vortex. Then its concentration was adjusted to  $1.5 \times 10^8$  CFU/mL based on a standard 0.5 McFarland [13].

### 3.6. Antimicrobial Susceptibility Test

The ability of the various extracts to inhibit growth of clinical bacteria isolates was determined using the Agar disc diffusion method sterile filter paper discs; 7 mm in diameter were impregnated with each extract concentration and dried at 30°C in the static incubator. They were then carefully placed aseptically with a forceps on the surface of the Mueller Hinton agar (DEFCO Laboratories, USA), plates that were preinoculated with the 24 hours culture of bacteria and 0.1 mL suspension ( $1.5 \times 10^8$  CFU/mL) of the bacteria isolates, respectively. The control antibiotics disc containing gentamycin solution was placed on each of the inoculated plates of Mueller Hinton agar. The plates were left on the bench undisturbed for few minutes, after which the bacterial plates were incubated at 37°C for 24 hours. The external diameters of visible zones of growth inhibition were measured after incubation [14, 15].

### 3.7. Minimum Inhibitory Concentration (MIC)

The minimum inhibitory concentration of extracts were determined by an agar dilution method according to the national committee for clinical laboratory standard (NCCLS) guidelines using a multipoint replicator and delivering 0.3 mL of standardized microbial suspension. The final concentration of extracts in the medium ranging was from 2 mg/mL to 256 mg/mL. The plates were incubated at 37°C for 18 - 24 hours and the MIC was defined as the lowest concentration of extracts inhibiting the visible growth. All determinations were performed in duplicate and growth control consisting of MH medium was included [16].

### 3.8. Minimum Bactericidal Concentration (MBC)

Cells from the plate showing no growth were sub cultured on nutrient agar plates to determine whether the inhibition was reversible or permanent. MBC was determined as the highest dilution (lowest concentration) at which no growth occurred on the plates [17].

### 3.9. Statistical Analysis

Average results were reported as the mean and standard error (mean  $\pm$  SE) and SPSS-18 statistical software, one way ANOVA followed by Turkey's test were used to do intergroup comparison, while considering  $P \leq 0.05$  as the significance level.

## 4. Results

This study reports the antimicrobial activity of 4 concentrations of *M. communis* against *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri*. The results antimicrobial effects of methanolic and aqueous *M. communis* extracts, by "the agar diffusion method" were presented in Tables 1 and Figure 1. In this study, aqueous extract showed no inhibition against *E. coli* and *S. flexneri* bacteria tested at 5 mg/mL concentrations.

## 5. Discussion

The diameter of inhibition zone in methanol extract is higher than aqueous extract. On the contrary, statistical analysis using Turkey's test analysis showed that there are no significant differences between methanol extract and aqueous extract used. The discs containing 20 mg/mL of methanolic extract showed the highest mean zones of Inhibition against *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri*, while the discs containing 5 mg/mL showed the lowest inhibitory zones. Methanolic extract of *M. communis* showed the highest mean zones of inhibition ( $20 \pm 0.28$  mm) at 20.0 mg/mL on *S. epidermidis*. Aqueous *M. communis* extract showed the highest mean zones of inhibition ( $18 \pm 0.28$  mm) at 20 mg/mL on *S. epidermidis*.

Turkey's test range tests showed significant difference among the mean diameter inhibition zones for the ethanolic stem extracts of *M. communis*. The findings in this study pointed out that the higher the concentrations of the extracts, the higher the sensitivities of *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri* to the methanolic extract of *M. communis* as evidenced by the increased size of the bacterial growth inhibition zones. Since ancient civilization, natural sources especially plants are used as medicinal therapy because they contain several components which

are believed to cure various infectious diseases. The biodiversity of plants provides an important source of chemical compounds, which have much therapeutic application such as antiviral, antibacterial, antifungal and anticancer activities [18]. Therefore, the aim of this study was to investigate the possible inhibitory activity of *M. communis* extracts against pathogenic strains causing infection.

Thus, exhibiting concentration dependent activity and these results are in conformity with Chandrasekaran et al. [19] and Chomnawang et al. [20]. Iauket al. [21] also showed that the higher the concentrations of the *Vernonia amygdalina*, the larger the diameter of the bacterial growth inhibition zones.

The MIC of methanolic extract of *M. communis* for *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri* were 2, 4, 16 and 32 mg/mL, respectively. However, MIC of the aqueous extract of *M. communis* for *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri* were 4, 16, 64 and 128 mg/mL, respectively. The mechanism of action of extract plant and essential oil and their components as antimicrobials has not been fully elucidated. This is complicated by the fact that there are a large number of chemical compounds present in extract and EOs and often they are all needed for antibacterial activity and the extract and EOs does not seem to have a specific cellular target. Thus the antimicrobial mechanism of extract and EOs may not be attributable to one specific mechanism, but rather there may be several targets in the cell. Most of the focus on antimicrobial mechanisms for extract and EOs has been on the cell membrane and targets interconnected with the membrane. For bioactivity, the extract and EOs pass through the cell wall and cytoplasmic membrane [22].

The MBC of methanolic extract of *M. communis* for *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri* were 4, 8, 32 and 64 mg/mL, respectively. But MBC of aqueous extract of *M. communis* for *S. epidermidis*, *E. faecalis*, *E. coli* and *S. flexneri* were 8, 32, 128 and 256 mg/mL, respectively. In general, among the tested microbial strains, *S. epidermidis* were found to be more sensitive to many of the test agents than *E. faecalis*, *E. coli* and *S. flexneri*.

The antibacterial activity was more pronounced on the Gram (+) bacteria (*S. epidermidis* and *E. faecalis*) than the Gram (-) bacteria (*E. coli* and *S. flexneri*). The reason for the difference in sensitivity between Gram (+) and Gram (-) bacteria might be ascribed to the differences in morphological constitutions between these microorganisms, Gram (-) bacteria having an outer phospholipidic membrane carrying the structural lipopolysaccharide components [23]. This makes the cell wall impermeable to antimicrobial chemical substances. The Gram (+) bacteria on the other hand, is more susceptible having only an outer peptidoglycan layer which is not an effective permeabil-

**Table 1.** Average Diameter (mm) of Microbial Free Zone Area of Aqueous and Methanolic *Myrtus communis* Extracts Concentrations on *Staphylococcus epidermidis*, *Enterococcus faecalis*, *Escherichia coli* and *Shigella flexneri* (Disk Agar Diffusion Method)

Type of Extract	The Concentration of <i>Myrtus communis</i> Extracts, mg/mL <sup>a</sup>			
	5	10	15	20
<b>Aqueous, microorganism</b>				
<i>S. epidermidis</i>	9.00 ± 0.54	12.40 ± 0.50	16.00 ± 0.52	18.00 ± 0.28
<i>E. faecalis</i>	8.00 ± 0.52	11.10 ± 0.54	15.80 ± 0.54	17.30 ± 0.28
<i>E. coli</i>	-	8.20 ± 0.52	10.10 ± 0.28	12.30 ± 0.50
<i>S. flexneri</i>	-	7.20 ± 0.54	9.40 ± 0.28	12.00 ± 0.50
<b>Methanolic, microorganism</b>				
<i>S. epidermidis</i>	10.80 ± 0.58	14.00 ± 0.54	17.80 ± 0.54	20.0 ± 0.28
<i>E. faecalis</i>	9.90 ± 0.58	14.30 ± 0.50	16.40 ± 0.54	19.70 ± 0.28
<i>E. coli</i>	8.00 ± 0.54	10.00 ± 0.57	12.10 ± 0.54	14.00 ± 0.58
<i>S. flexneri</i>	7.70 ± 0.58	9.80 ± 0.54	12.00 ± 0.28	13.60 ± 0.57

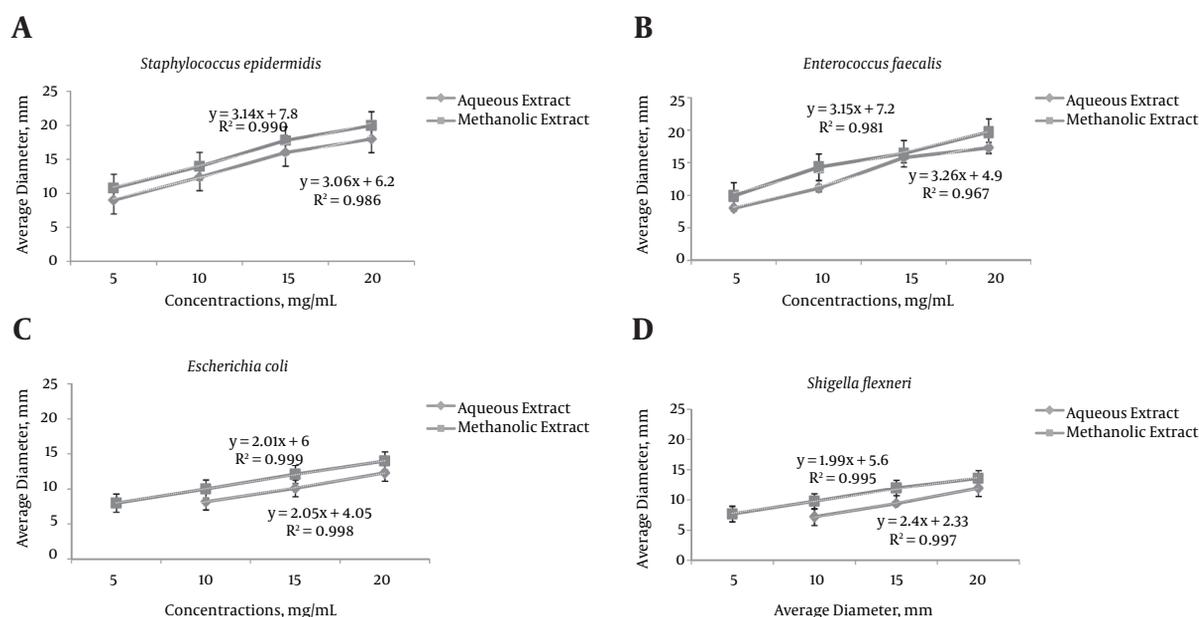
<sup>a</sup>Values are expressed as means ± SD; n = 3.**Table 2.** Minimum Inhibitory Concentration (MIC) of Aqueous and Ethanolic Extract of *Myrtus communis* on *Staphylococcus epidermidis*, *Enterococcus faecalis*, *Escherichia coli* and *Shigella flexneri*

Type of Extract	Concentration, mg/mL <sup>a</sup>									
	2	4	8	16	32	64	128	256	Negative	Positive
<b>Aqueous, bacteria species</b>										
<i>S. epidermidis</i>	+	+	-	-	-	-	-	-	-	+
<i>E. faecalis</i>	+	+	+	+	-	-	-	-	-	+
<i>E. coli</i>	+	+	+	+	+	+	-	-	-	+
<i>S. flexneri</i>	+	+	+	+	+	+	+	-	-	+
<i>S. epidermidis</i>	-	-	-	-	-	-	-	-	-	+
<i>S. flexneri</i>	+	-	-	-	-	-	-	-	-	+
<b>Methanolic</b>										
<i>S. epidermidis</i>	+	+	+	-	-	-	-	-	-	+
<i>E. faecalis</i>	+	+	+	+	-	-	-	-	-	+

<sup>a</sup>+, grow; -, not grow; n = 3.**Table 3.** Minimum Bactericidal Concentration (MBC) of Aqueous and Ethanolic Extract of *Myrtus communis* on *Staphylococcus epidermidis*, *Enterococcus faecalis*, *Escherichia coli* and *Shigella flexneri*

Type of Extract	Concentration, mg/mL <sup>a</sup>									
	2	4	8	16	32	64	128	256	Negative	Positive
<b>Aqueous, bacteria species</b>										
<i>S. epidermidis</i>	+	+	-	-	-	-	-	-	-	+
<i>E. faecalis</i>	+	+	+	-	-	-	-	-	-	+
<i>E. coli</i>	+	+	+	+	+	+	-	-	-	+
<i>S. flexneri</i>	+	+	+	+	+	-	-	-	-	+
<i>S. epidermidis</i>	+	-	-	-	-	-	-	-	-	+
<i>E. faecalis</i>	+	+	-	-	-	-	-	-	-	+
<b>Methanolic</b>										
<i>E. coli</i>	+	+	+	+	-	-	-	-	-	+
<i>S. flexneri</i>	+	+	+	+	+	-	-	-	-	+

<sup>a</sup>+, grow; -, not grow; n = 3.



**Figure 1.** Antimicrobial Activity of Methanolic and Aqueous at Different Concentration of *Myrtus communis* Leaves Extracts on A, *Staphylococcus epidermidis*, B, *Enterococcus faecalis*, C, *Escherichia coli* and D, *Shigella flexneri*

ity barrier. Therefore, the cell walls of Gram (-) organisms which are more complex than the Gram (+) ones act as a diffusional barrier and making them less susceptible to the antimicrobial agents than are Gram (+) bacteria. In spite of this permeability differences, however, some of the extracts have still exerted some degree of inhibition against Gram (-) organisms as well [24].

Rasooli et al. [25] reported the major components *M. communis* were  $\alpha$ -pinene (29.4%), limonene (21.2%), 1, 8-cineole (18%), linalool (10.6%), linalyl acetate (4.6%) and  $\alpha$ -terpineole (3.1%). This study favors the report that the essential oils with high monoterpenes hydrocarbons are very active against microorganisms. These secondary metabolites exert antimicrobial activity through different mechanisms. Tannins form irreversible complexes with proline rich protein, resulting in the inhibition of cell protein synthesis, flavonoids complex with extracellular-soluble proteins and bacterial cell wall proteins, while lipophilic flavonoids disrupt microbial cells membranes [26].

Mert et al. [27] investigated the antimicrobial activities of n-hexane, methanol, ethanol, ethyl acetate and water extracts of *M. communis* L. leaves against *E. coli*, *S. aureus*, *S. epidermidis*, *Salmonella typhimurium*, *Enterobacter cloacae*, *E. faecalis* and *P. aeruginosa* as bacteria and *Candida albicans* as yeast like fungi by disc diffusion method. Results

showed all the extracts inhibited the growth of *E. coli*, *S. epidermidis*, *S. typhimurium* and *P. aeruginosa*. The growth of *E. coli* was only inhibited by the methanol extract. None of the tested extracts showed activity against *E. cloacae*, *E. faecalis* and *C. albicans*.

On the basis of the above, results showed that methanol extract of *M. communis* exhibited a greater inhibition compared with aqueous extract. Alizadeh-Behbahani et al. [28] reported that the most of the antimicrobial active compounds were soluble in polar solvent such as ethanol instead of water. Koffi-Nevry et al. [29] the effect of *Capsicum annum* and *Capsicum frutescens* methanol and aqueous extracts on selected bacteria (*S. aureus*, *S. typhimurium*, *Vibrio cholerae*, *Pseudomonas aeruginosa*, *E. coli*, and *S. dysenteriae*) were investigated. Both extracts were found to be effective against *V. cholerae*, *S. aureus* and *S. typhimurium*, while methanol extracts showed the greatest effect. The extract from *Capsicum annum* showed a higher antibacterial activity than the one from *Capsicum frutescens*. The MIC of methanol and aqueous extracts were 0.2 mg/mL and 0.25 mg/mL, respectively. MBC values of both extracts ranged from 1 to 2.5 mg/mL, this result is consistent with the findings of this study. This was also reported by Parekh et al. [30], the aqueous and ethanolic extracts of *Launaea procumbens* Roxb. (Labiatae), *Vitis vinifera* L. (Vitaceae) and *Cyperus rotundus* L. (Cyper-

aceae) were evaluated for antimicrobial activity against clinically important bacteria viz. Ethanollic extracts were more potent than aqueous extracts and activity were concentration dependent, this result is consistent with the findings of this study.

In this study, a limit of our research is the small amount of the extract that did not allow performing the MIC towards all the strains used. These data are encouraging even if further additional ‘in vitro’ testing and large clinical studies are necessary to verify the potential use of the extract of *M. communis* as antibacterial drug. Given the excellent results obtained in this study, we would expand the research with further studies, to value the possible cytotoxic effects, and eventually to perform tests using ‘in vivo’ mouse model.

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### Footnotes

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